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Synthesis and Study of Calix[6]cryptamides: A New Class of Heteroditopic Receptors that Display Versatile Host–Guest Properties Toward Neutral Species and Organic Associated Ion-Pair Salts

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Abstract: The synthesis of a new family of molecular receptors, namely the calix[6]cryptamides, was achieved through an original [1+1] macrocyclization step that consists of a peptidecoupling reaction between tripodal triscarboxylic acids and a calix[6]trisamine subunit. Several C_3 - or C_3 -symmetrical calix[6]arene-based compounds capped by a trisamido cryptand unit on the narrow rim have been obtained, with the more flexible partners leading to the best yields. These calix[6]cryptamides exhibit two favorably positioned

Introduction

There is a growing interest in the design of synthetic receptors that are able to recognize neutral molecules or charged species through specific interactions.^[1] These receptors can find potential applications in many areas, such as sensing, modeling of enzymic active sites, catalysis, nanoscience, drug delivery and separation science. Readily available calixarenes have emerged as very attractive building blocks for the development of such host–guest systems.[2] Of the different

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binding sites for the complexation of organic-associated ion pairs in close proximity: a well-defined calix[6]arene cavity suitable for the inclusion of ammonium ions and a cryptamide unit for the coordination of anions. We demonstrate one example, chiral calix[6]cryptamide 12, that constitutes a heteroditopic receptor capable of cooperatively

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binding both a primary ammonium ion and its chloride counterion, thanks to a combination of polarization and induced-fit effects. In addition, the hydrophobic calixarene cavity of 12 can strongly bind neutral guests through hydrogen bonding and is capable of discriminating between different enantiomers. All these versatile host–guest properties differ greatly from those observed in the parent calix[6]azacrypt-

oligomers, calix[4]arenes have been extensively studied, since control of their conformational mobility and their selective functionalization can be more easily achieved. However, calix[4]arenes suffer from the smallness of their hydrophobic cavity and consequently they have mostly been used as molecular platforms for the preorganization of a binding site outside the cavity.^[2,3] The larger calix[6]arenes display a cavity size that is well adapted for the inclusion of organic guests $[4]$, but usually their high conformational flexibility needs to be restricted first. To do this, different strategies such as self-assembly,^[5] metal-ion coordination,^[6] and grafting of covalent bridges have been developed.[7] We have synthesized calix[6]arenes bearing a tripodal azacryptand cap on the narrow rim.[8] These calix[6]azacryptands exhibit remarkable host–guest properties toward polar neutral molecules or cationic species (ammonium and metal ions), thanks to the presence of the basic azacryptand cap that preorganizes the cavity and provides a tunable binding site.^[9] In the course of designing versatile calixarene based endo-receptors for organic guests, we wanted to test the feasibility of grafting a tripodal amido cap (that is, a cryptamide unit) onto a calix[6]arene core. So far, only calix[6]arenes that have intramolecular dipodal amido bridges have been described.^[10,11] Apart from the synthetic challenge, the

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target calix[6]cryptamides were particularly attractive, since they were predicted to possess both an ammonium ion binding site in the form of the hydrophobic cavity, and an anion binding site in the form of the tripodal arrangement of the amide groups.^[12] Neutral heteroditopic receptors capable of simultaneously binding cations and anions are currently being intensively studied since they present the advantage of avoiding the competitive ion-pairing of the guest salt.^[13] In this regard, heteroditopic calix[4]arene-based receptors

that possess amido or urea groups have been already produced.[14] However, owing to the smallness of their cavity they are mostly used for the complexation of metal ions and their counterions.[15] With the larger calix[5,6]arenes, the introduction of urea moieties led to receptors capable of recognizing organic ion-pair salts, but only a few examples have been reported so far. $[16-18]$

Here we describe an efficient synthesis of calix[6]cryptamides, a new class of bifunctional receptors, and also the NMR spectroscopic host–guest studies of chiral calix[6]cryptamide 12. This versatile heteroditopic receptor exhibits unique host properties towards both organic-associated ion pair salts and neutral guests.

Results and Discussion

Synthesis of calix[6]cryptamide 4: For the key step of the synthesis of calix[6]cryptamides, we chose a [1+1] mac-

rocyclization reaction between two tripodal subunits, since this strategy has proved its efficiency with the parent cal $ix[6]$ azacryptands.^[8] The reaction conditions for the macrocyclization step were first tested and optimized with calix[6] trisamine 1 and well-preorganized tris-electrophilic partners 2 and 3 derived from a cyclotriveratrylene (CTV) skeleton. In addition, we were interested in producing a multitopic receptor that displayed two concave subunits linked by a trisamido binding site.^[19]

The preparation of calix[6]trisamine 1 from the C_{3v} symmetrical 1,3,5-tris-O-methylated tBu-calix[6]arene has already been reported by us. The synthesis is an efficient three-step sequence: per-alkylation with ethylbromoacetate in presence of a strong base (NaH), reaction with ammonia in MeOH, and subsequent reduction of the obtained amide groups by BH₃/THF (79% overall yield).^[8g] (\pm)-CTV triscarboxylic acid derivative 2 was synthesized according to the procedure described in the literature.[20]

First, the [1+1] macrocyclization reaction between calixarene 1 and trisacyl chloride 3 was tested under classical high dilution conditions (trisacyl chloride 3 was prepared quantitatively from CTV 2 and oxalyl chloride in dichloromethane).^[21] The desired calix[6]cryptamide 4 (15%) was isolated after purification by flash chromatography (FC) (Scheme 1, entry 1 of Table 1).

Scheme 1. Synthesis of calix[6]cryptamide 4; i) $(COCl)_2$, CH_2Cl_2 ; ii) see Table 1.

Table 1. Synthesis of calix[6]cryptamide 4 through reaction between calix[6]trisamine 1 and CTV derivatives 2 or 3.

Entrv	CTV derivative	Solvent	$Conc.[M]^{[a]}$	Reaction conditions ^[b]	Yield[%][c]
		toluene	2.6×10^{-3}	TEA, 0° C then RT, 15 h	15
γ		$CHCl3/CH3NO2; 4:1$	1.0×10^{-2}	PyBOP, 50°C, 2 h	46
		$CHCl3/CH3NO2; 4:1$	1.0×10^{-2}	HBTU, 50 °C, 2 h	42
		$CHCl3/DMF$; 2:1	1.0×10^{-2}	HBTU, 50 °C, 2 h	41
		$CHCl3/CH3NO2; 4:1$	1.0×10^{-1}	PyBOP, 50°C, 15 h	10
		$CHCl3/CH3NO2; 4:1$	4.0×10^{-4}	HBTU, 50 °C, 15 h	$<$ 5[d]

[a] Concentration of 1 and CTV derivative. [b] Reactions with HBTU and PyBOP were performed by the slow addition of a mixture of the coupling reagent and TEA (6 equiv each; see the Experimental Section). [c] The yields correspond to the isolated compound 4 after purification by flash chromatography. [d] This yield was estimated by using an NMR spectrum of the crude reaction mixture.

> With the aim of improving the yield of the macrocyclization reaction, we decided to test the direct reaction between calix[6]trisamine 1 and CTV-trisacid 2 in the presence of peptide-coupling reagents such as HBTU or PyBOP.[22] Mixtures of solvents such as $CHCl₃/CH₃NO₂$ or $CHCl₃/DMF$ were used to solubilize the two tripodal partners and the coupling reagent.[23] In addition, the influence of the concentration of subunits 1 and 2 was studied (entries 2–6, Table 1). The highest yield for 4 (46%) was obtained when subunits 1 and 2 were reacted at 50° C and at a concentration of 10^{-2} M in a 4:1 mixture of CHCl₃/CH₃NO₂, with PyBOP as the coupling reagent (entry 2, Table 1).[24] It is worth noting that the nature of the coupling reagent and the solvent have little influence on the yield (41–46%) (entries 3 vs. 2 and 4 vs. 3, respectively, Table 1). In contrast, high concentration (10^{-1}m) or high dilution $(4 \ 10^{-4} \text{m})$ of the

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two subunits led to low yields (entries 5 and 6, Table 1). The ¹H NMR spectra of the crude reaction mixture show in both cases the presence of multiple species displaying ill-defined resonances.[25]

This first set of experiments shows that the peptide-coupling reaction can constitute an efficient strategy for [1+1]

macrocyclization reactions between tripodal units^[26] and thus we decided to test these reaction conditions for the synthesis of various calix[6]cryptamides.

Synthesis of calix[6]cryptamides 10–13: Calix[6]trisamine 1 was reacted with triscarboxylic acid derivatives 5–9, which display different degrees of flexibility^[27] (Scheme 2). All the reactions were performed under the optimal conditions determined for 2 (50 $^{\circ}$ C, 6 equiv of TEA, 6 equiv of coupling reagent, 10^{-2} M in a 4:1 mixture of CHCl₃/CH₃NO₂ or $CHCl₃/DMF$; Table 2). The choice of coupling reagent was governed by purification considerations. The reactions were monitored by ¹H NMR spectroscopy and the obtained calix[6]cryptamides were purified by flash chromatography. In the case of triscarboxylic acid 5, a complex mixture of calixarene derivatives that displayed broad NMR resonances was obtained. However, the use of triscarboxylic acids 6–9 led to the corresponding calix[6]cryptamides 10–13 in moderate to

[a] The yields correspond to the isolated compounds after purification by flash chromatography. [b] The yield was calculated while taking into account the presence of about 1 equiv of 1,1,3,3-tetramethylurea in the final product (¹H NMR estimation).

> good yields (13–58%, Table 2, entries 2–5). Surprisingly, the efficiency of the macrocyclization reaction augments with increasing flexibility of the tripodal carboxylic acid. This can be rationalized if one considers that the bulkiness of the intermediate benzotriazole ester groups disfavors the [1+1] macrocyclization pathway in the case of rigid partners.

> All these results show that the use of the peptide-coupling reaction for the [1+1] macrocyclization step can be extended to structurally different tripodal partners and gives access to original calixarene based compounds that are either closed at the narrow rim by a grid-like amido cap (4, 10–12) or possessing a calix[6]tube skeleton (13).

> NMR spectroscopic study of the conformational properties of calix[6]cryptamides 4 and 10–13: The conformational properties of calix[6]cryptamides 4 and 10–13 were investigated through extensive NMR studies in CDCl₃. All the resonances of the ¹ H NMR spectra were attributed by means

Scheme 2. Syntheses of calix[6]cryptamides 10–13; see Table 2 for the reaction conditions. The calix[6]cryptamides are shown in their major conformation, which has been deduced from NMR spectroscopic studies (see below).

of 2D NMR experiments (COSY, HMQC, HMBC). Due to their C_3 (compound 4) or C_{3v} (compounds 10, 11, and 13) symmetry, calix[6]cryptamides display simple ¹H NMR patterns. In the case of 12, a minor dissymmetric conformer with a *t*Bu group included in the calixarene cavity was observed; however, recording the spectrum at 330 K in a 3:1 mixture of $CDCl₃/CD₃OD$ led to a unique $C₃$ -symmetrical NMR signature (see the Supporting Information). The minor conformer observed in CDCl₃ is probably due to a dissymmetrical arrangement of the cap, induced by an intramolecular hydrogen-bonding network between the amido arms. Interestingly, the chirality of the CTV cap of 4 and 12 is transmitted to the calixarene core as shown by the diastereotopy of the ArCH₂ and ArH protons (see Figure 1a for 4). It is worth noting that in the case of the tail-to-tail biscalix[6]arene 13, both calixarene frameworks are differentiated as attested to by the two sets of signals displayed by the OMe groups and the $ArCH₂$ protons (Figure 1b).

Typical ¹H resonances of the calixarene subunits of cal $ix[6]$ cryptamides 4 and 10–13 at 294 K are summarized in Table 3. From these NMR spectroscopic data, it is possible to deduce the following conformational features:

1) For all the calix[6]cryptamides, the $ArCH₂$ protons of the calixarene moiety display well-defined differentiated signals and, when the spectra are recorded at high temperature (330 K), these resonances are not affected. This

Figure 1. ¹H NMR spectra (CDCl₃, 294 K) of: a) calix[6]cryptamide 4; b) calix[6]cryptamide 13. S: solvent (CHCl3); W: water; "cal" and "CTV" represent the calixarene and cyclotriveratrylene subunits, respectively.

 δ (ppm)

Table 3. Selected chemical shifts [ppm] of the calixarene subunit protons of calix[6]cryptamides 4 and 10–13 $(CDCl_3, 294 K).$

Entry	Compound	O _{NHCO}	$\Delta\delta$ _{ArH}	$\Delta\delta_{t\mathrm{Bu}}$	O_{ArCH2ax}	O_{ArCH2ea}	$\sigma_{\rm OMe}$
	$\mathbf{4}^{[\mathrm{a}]}$	7.31	0.59	0.52	4.33	3.39	2.10
	10	6.67	0.31	0.36	4.37	3.34	2.24
		9.11	0.63	0.63	4.58	3.38	1.66
	$12^{[a,b]}$		0.32	0.35	4.43	3.45	2.60
	13	7.81	0.61/0.62	0.55/0.57	4.55/4.60	3.44/3.45	2.45/2.48

[a] Average value due to the diastereotopy. [b] The ${}^{1}H NMR$ spectrum was recorded at 330 K in CDCl₃/ $CD₃OD (3:1).$

clearly indicates an inhibition of the cone–cone inversion of the calixarene core due to the presence of the tripodal covalent cap on the narrow rim.

- 2) The large $\Delta \delta_{A r H}$ and $\Delta \delta_{t B u}$ values of 4, 11, and 13 indicate that their calixarene subunit adopts a flattened cone conformation, whereas a straighter cone is observed for 10 and 12 (see the structures displayed in Schemes 1 and 2).[28]
- 3) In compounds 4, 11, and 13, which have a calixarene core in a flattened cone conformation, the highfield resonances of the OMe groups is due to their location in the calixarene cavity (Table 3, entries 1, 3, and 5). The methoxy groups of 12 are less directed toward the inside of the cavity since these compounds display a straighter conformation (Table 3, entry 4). In the case of 10, the close proximity of the aromatic rings of the cap should be responsible, to some extent, for the high-field shift of the methoxy groups (Table 3, entry 2).
- 4) Compared with all the other calix[6]cryptamides, compound 11 displays a strong downfield resonance for the NHCO protons (Table 3, entry 3). This clearly indicates the presence of stable six-membered intramolecular hydrogen-bonded rings with the oxygen atoms in ortho positions (see Scheme 2).

This NMR spectroscopic conformational study shows that, thanks to their covalent tripodal cap, calix[6]cryptamides

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possess a well-defined cavity ideally preorganized for host– guest applications. Binding studies of charged or neutral species were conducted with each calix[6]cryptamide and results concerning the unique host–guest properties of calix[6]cryptamide 12 are described below.

Study of the host–guest properties of calix[6]cryptamide 12:First, the host–guest properties of calix[6]cryptamide 12 toward neutral molecules G were evaluated by 1 H NMR spectroscopy. No inclusion was observed, even at low temperature (263 K), upon the addition of an excess $(> 20$ equiv) of EtOH, DMSO, or apolar molecules such as CH₂Cl₂ or $Et₂O$ to a solution of 12 in CDCl3. In contrast, calix[6] cryptamide 12 was found to be a remarkably efficient host for the endo-complexation of amide or urea type guests such as pyrrolidin-2-one (PYD),

imidazolidin-2-one (IMI) or (\pm)-4-methylimidazolidin-2-
one (MIMI)^[29] (Scheme 3). one $(MIMI)^{[29]}$ Indeed, in each case the addition of a few equivalents (2 to 10 equiv) of these polar molecules (G) led to new C_3 -symmetrical NMR spectroscopic patterns (Figure 2a, b for $G=$ IMI at 263 K) displaying highfield signals, which have been assigned to the included guests through NOESY experiments (see the Supporting Information). These NMR data show unambiguously that the new species correspond to the host– guest complexes 12 OG. In all cases, the in and out exchange process of the guest within the cavity of 12 was slower than the NMR spectroscopic timescale. The NMR complexationinduced upfield shifts (CIS, see Table 4, entries 1–3) were quasi-similar to those obtained with the closely related protonated calix[6]azacryptands.[8d,g]

Moreover, in the case of IMI, Scheme 3. Host–guest properties of calix[6]cryptamide 12 toward neutral or charged species in CDCl₃.

Figure 2. ¹H NMR spectra (CDCl₃, 263 K): a) **12**; b) **12**+2.6 equiv of IMI; c) **12**+15 equiv of PrNH₃⁺Cl⁻. \circ : IMI (free); \bullet : IMI (in); \triangledown : PrNH₃⁺ (free), \bullet : PrNH₃⁺ (in). S: solvent (CHCl₃); W: water; "cal" and "CTV" represent the calixarene and cyclotriveratrylene subunits respectively.

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Table 4. Complexation-induced upfield shifts (CIS) and association constants (K) of the guests G and $RNH_3^+Cl^-$ in the case of $12\supset G$ and $12\supset RNH_3^+Cl^-$.

		$\text{CIS}[ppm]^{[\text{b}]}$				
Entry	G or $RNH3$ ⁺ $Cl-$	$K\,[\mathrm{M}^{-1}]^{[\mathrm{a}]}$	α	β	γ	δ
$\mathbf{1}$	δ .α \star NΗ γ β	$\operatorname{nd}^{[\operatorname{c}]}$			$-3.43_{\text{(CH2N)}}^{[d,e]}$ $-2.71_{\text{(CHN)}}^{[d]}$ $-3.60_{\text{(CH2N)}}^{[f,e]}$ $-2.71_{\text{(CHN)}}$ ^[f]	$-3.16^{[d]}$ $-3.26^{[f]}$
\overline{c}	Hβ \checkmark α γ	250		$-0.81_{\rm (CH2CO)}$ $+0.74_{(NH)}$	$-3.82_{(CH2)}$ $-3.32_{\text{(CH2N)}}$	
3	β_{H} α γ $NH \beta$	12800			-3.47	
$\overline{4}$	α β $NH3+Cl1$	$nd^{[c]}$	$nd^{[c]}$	-2.92		
5	NH_3 ⁺ Cl ⁺ α γ	230	-1.20	-3.09	-3.07	

[a] K calculated at 263 K and defined as: $K = \left[\frac{12 \cdot 10}{12}\times10^{-11}\right] \times 10^{-11} \text{ s}^{-1}$ ($\left[\frac{12 \cdot 10^{-11}}{200}\right] \times 10^{-11} \text{ s}^{-1}$ ($\left[\frac{12 \cdot 10^{-11}}{200}\right] \times 10^{-11} \text{ s}^{-1}$ errors estimated $\pm 15\%$. [b] CIS calculated at 263 K and defined as $\Delta\hat{\sigma} = \hat{\sigma}_{\text{complexed G}} - \hat{\sigma}_{\text{free G}}$ [c] nd: not determined. [d] Major diastereomer. [e] Average value of the diastereotopic protons. [f] Minor diastereomer.

NOE effects were observed between the methylene protons of the guest and the ArH protons of the aromatic rings that are directed toward the inside of the calixarene cavity (see the Supporting Information).[30] These results demonstrate that the neutral guests are accommodated in the calixarene cavity and not in the CTV one. Interestingly, the host undergoes a significant conformational rearrangement upon complexation; the calixarene subunit adopts a flattened cone conformation ($\Delta\delta_{\text{A}H}$ and $\Delta\delta_{\text{B}u} \geq 0.6$ ppm) with the methoxy groups expelled from the cavity ($\delta_{\text{OMe}} \geq 3.7$ ppm). Such an induced-fit process has already been evidenced with one of the parent calix[6]azacryptands.[8g] A significant downfield shift of the NHCO resonances of the calixarene upon inclusion of the guests G ($\Delta\delta_{\text{NHCO(cal)}}$) \geq 1.2 ppm, see Figure 2a, b for $G=IMI$ at 263 K) indicates a stabilization of the guests through hydrogen-bonding. In addition, as shown by the positive CIS of the NH of the included PYD (entry 2, Table 4), the guests are probably stabilized through strong hydrogen-bonding interactions with the phenolic oxygen atoms of the host (see structure displayed in Figure 2 for $G=IMI$). These observations rationalize the specific recognition by receptor 12 of neutral molecules that possess hydrogen-bond donor and acceptor groups such as amides and ureas. In strong contrast with all previous host–guest calix[6]arene-based systems, the polarization of the calixarene cavity through metal-ion complexation[9a] or protonation[9b] is not a requirement for the efficient endo-complexation of neutral guests by a calix[6]cryptamide host. In other words, these results constitute the first example of intracavity recognition of neutral molecules by a neutral calix[6]arenebased receptor.

Interestingly, we also found that C_3 symmetrical calix[6]cryptamide host 12 is able to perform intracavity chiral recognition. In fact, the host–guest study undertaken with (\pm) -

MIMI indicated the formation of two diastereomeric endocomplexes. A diastereomeric excess of 44% was determined at 263 K through integration of well-resolved signals corresponding to the included MIMI guest of each diastereomer (Figure 3). This remarkable result constitutes a rare and leading example of enantioselective discrimination in the heart of the cavity of a chiral calixarene based recep- $\text{tor.}^{\text{[5d-g,8d]}}$

In a second set of NMR spectroscopic experiments, the ability of calix[6]cryptamide 12 accommodate organic charged species was investigated at 263 K. Addition of an excess (up to 15 equiv) of ammonium salts $RNH_3^+Cl^-$ (R = Et or Pr) to a solution of 12 in

 $CDCl₃$ produced the corresponding *endo-complexes* $12\supset$ RNH₃⁺Cl⁻ (Scheme 3) with, in both cases, high-field signals characteristic of the presence of the alkyl chain of the ammonium ions in the heart of the calixarene cavity (Figure 2c for $R=Pr$).^[31] As observed with neutral guests, the *in* and out guest exchange was slower than the NMR timescale, and involved an induced fit process with the expulsion of methoxy groups from the calixarene cavity. As expected, it was shown that 12 behaves as a heteroditopic receptor, since in addition to the endo-complexation of the ammonium ion $RNH₃⁺$, the simultaneous binding of the chloride counterion by the amide groups of the host was clearly evidenced. Indeed, the resonances of the NHCO protons close to the calixarene subunit experienced a significant downfield shift (> 0.5 ppm at 263 K, see Figure 2c for R=Pr) indicating a strong hydrogen-bonding interaction with the counter anion. Moreover, the CIS of the CH₂N protons of the $PrNH₃⁺$ guest was found to be abnormally low compared to those at the β or γ position of the ammonium group (Table 4,

Figure 3. High-field region of the 1 H NMR spectrum (CDCl₃, 263 K) of 12 MIMI. "M" and "m" represent "major diastereomer" and "minor diastereomer", respectively.

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entry 5). This is in contrast with previously reported host– guest systems involving calix[6]arene-based receptors and ammonium ions, which all display high CIS for protons at the α , β , or γ position of the ammonium group (that is, -2.0 to -3.2 ppm). These NMR data are compatible with the specific coordination of the chloride anion in close proximity to the cationic ammonium group, through a convergent arrangement of hydrogen-bonding NH groups (see Figure 2c). Concerning the ammonium ion, in addition to the electrostatic interaction with its counterion, it is probably stabilized through a combination of CH- π interactions and hydrogen bonding to the ethereal oxygen atoms that have the amido arms, as this has been already observed on several XRD structures of closely related endo-complexes.^[5f,8b] Interestingly, the use of low-coordinating counterions (picrate and tetrabutylammonium) demonstrated the remarkable cooperative binding of the associated ion pair. When a large excess (about 30 equiv) of $nBu_4N^+Cl^-$ was added to a solution of 12 in CDCl₃, the NMR spectrum of the receptor remained quasi-unchanged and only a tiny downfield perturbation of its NHCO protons was observed $(\Delta\delta=0.06$ ppm at 263 K). On the other hand, when the bulkier and less densely charged picrate anion was used in place of the chloride anion, no inclusion of the propylammonium ion was detected even after a prolonged time.^[32] Thus, the complexation of the chloride anion can only proceed when an ammonium ion is present in the calixarene cavity and conversely, without Cl⁻, host 12 is inefficient at binding the ammonium ion. This remarkable positive cooperativity is probably due to both structural and electronic alterations of the receptor: on one hand each partner of the associated ion pair polarizes the receptor and on the other hand, the anion coordination contributes to the preorganization of the binding site of its counterion through an induced fit process. Indeed, the binding of Cl⁻ induces the expulsion of the OMe groups from the cavity and brings the oxygen atoms of the ammonium binding site closer. Only a few examples of such metal-free allosteric controls of the complexation of an associated ionpair salt have been reported.^[15,18,33]

Lastly, it was also possible to estimate the association constants (K) of PYD, IMI, and PrNH₃⁺Cl⁻ for the receptor 12 (Table 4) from the ratio of the peak intensities of the host– guest complex and free partners.^[34] A strong preference was found for IMI with respect to the two other guests. This emphasizes the remarkable complementary fit between IMI and the calixarene core in terms of size, shape, and electronic structure.[35] As a result, it was possible to release the ammonium salt quantitatively from the endo-complex $12\supset PrNH_3^+Cl^-$ by the addition of a slight excess of IMI, resulting in the formation of the *endo-complex* 12 JMI (Scheme 3).

Conclusion

In summary, we have shown that the peptide-coupling reaction constitutes a valuable strategy for [1+1] macrocyclization reactions between tripodal partners. The obtained C_3 or C_{3v} -symmetrical calix[6]cryptamides exhibit two distinct well-preorganized binding sites in close proximity. As revealed through NMR spectroscopic studies, the chiral calix[6]cryptamide 12 behaves as a heteroditopic receptor with unique host–guest properties, since it can accommodate neutral molecules in the heart of the calixarene cavity, perform intracavity chiral recognition, or cooperatively bind organicassociated ion-paired salts. These results contrast with the host–guest properties of the parent calix[6]azacryptands and constitute a remarkable example of the synergistic combination of a polyamide site and a calix[6]arene structure. Current work is directed toward studying the host–guest properties of this promising class of molecular receptors, with a focus on the design of multitopic anion sensors.

Experimental Section

General procedures: All reactions were performed under an inert atmosphere. CH₂Cl₂ and CH₂NO₂ were distilled over CaH₂ under argon. Toluene and CHCl₃ were distilled over P_2O_5 under argon. DMF was distilled over a mixture of $MgSO₄$ and silica gel under argon. Silica gel (230– 400 mesh) was used for flash chromatography purifications. ¹H and 13C NMR spectra were recorded at 300 and 75 MHz, respectively. Chemical shifts are expressed in ppm. Traces of residual solvent or poly(dimethylsiloxane) were used as internal standard. All the ¹H NMR spectra signals were attributed through 2D NMR analyses (COSY, HMQC, HMBC).

Calix[6]cryptamide $4Method A$: In a sealed tube, oxalyl chloride (100 µL, 1.153 mmol) was added to CTV derivative 2 (67 mg, 0.115 mmol) in anhydrous CH₂Cl₂ (2 mL). The mixture was heated for 3 h at 50 $\rm{^{\circ}C}$ (a clear solution was obtained after about 30 min), then concentrated under reduced pressure yielding CTV derivative 3 as a white solid.

At 0° C, a solution of crude compound 3 (0.115 mmol) in anhydrous toluene (10 mL) and a solution of calix[6]trisamine 1 (120 mg, 0.105 mmol) and TEA $(54 \mu L, 0.388 \text{ mmol})$ in anhydrous toluene (10 mL) were simultaneously added to toluene (20 mL) over a period of 30 min. The mixture was stirred 15 h at room temperature and the solvent was then removed under reduced pressure. The residue was dissolved in CH_2Cl_2 (50 mL) and washed with an aqueous NaOH solution (1m, 20 mL). The aqueous layer was then extracted with CH_2Cl_2 (1 × 10 mL) and the combined organic layers were evaporated under reduced pressure. The crude residue was purified by flash chromatography on silica gel (CH₂Cl₂/AcOEt; 50:50), yielding calix[6]cryptamide 4 as a white solid (25 mg, 15%).

Method B: Anhydrous CHCl₃ (10 mL) and anhydrous CH₃NO₂ (2.5 mL) were added to calix[6]trisamine 1 (100 mg, 0.0874 mmol) and CTV derivative 2 (51 mg, 0.0874 mmol). The mixture was heated at 50 °C until a clear solution was obtained (about 2 h), then a solution of PyBOP (273 mg, 0.524 mmol) and TEA (75 μ L, 0.524 mmol) in anhydrous $CH₃NO₂$ (3 mL) was slowly added at 50 °C (0.5 mL every 30 min). The reaction mixture was stirred for 2 h at 50 $^{\circ}$ C and then the solvents were removed under reduced pressure. The residue was dissolved in CH_2Cl_2 (40 mL) and washed with H_2O (10 mL). The aqueous layer was then extracted with CH₂Cl₂ (2×5 mL) and the combined organic layers were evaporated under reduced pressure. The crude residue was purified by flash chromatography on silica gel (eluent: $CH_2Cl₂/ACOE$ t; 50:50), yielding calix[6]cryptamide 4 as a white solid (67 mg, 46%). M.p.: 210 °C (decomp); ¹H NMR (300 MHz, CDCl₃): δ 0.76 (s, 27 H; tBu), 1.27 (s_b, 27 H; tBu), 2.10 (s_b , 9 H; OMe_{cal}), 3.38 (d, J = 14 Hz, 3 H; ArC H_{eq} cal), 3.41 (d, J=15 Hz, 3H; ArC H_{eqcal} al), 3.57 (d, J=14 Hz, 3H; ArC H_{eq} CTV), 3.52– 3.69 (m, 3H; OCH₂CH₂N), 3.86 (s, 9H; OMe_{CTV}), 3.90 (d, $J=8$ Hz, 3H; OCH₂CH₂N), 3.90–4.03 (m, 3H; OCH₂CH₂N), 4.06 (d, $J=7$ Hz, 3H; OCH₂CH₂N), 4.30 (d, J = 15 Hz, 3H; ArCH_{axcal}), 4.37 (d, J = 15 Hz, 3H;

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ArCH_{axca}), 4.45 (d, J=15 Hz, 3H; OCH₂CO), 4.68 (d, J=15 Hz, 3H; OCH₂CO), 4.76 (d, J = 14 Hz, 3H; ArCH_{axCTV}), 6.59 (s, 3H; ArH_{cal}), 6.64 (s, 3H; Ar H_{cal}), 6.87 (s, 3H; Ar H_{CTV}), 6.88 (s, 3H; Ar H_{CTV}), 7.21 (s, 6H; ArH_{cal}), 7.31 ppm (s_b, 3H; NHCO); ¹³C NMR (75 MHz, CDCl₃) δ 30.1, 31.3, 31.6, 34.1, 34.3, 36.3, 40.2, 56.8, 60.7, 70.4, 71.9, 114.7, 117.1, 123.8, 124.0, 127.8, 128.1, 132.2, 132.8, 133.5, 134.3, 146.0(9), 146.1(5), 146.8, 148.9, 151.9, 154.7, 169.5 ppm; IR (KBr): $\tilde{v} = 3700 - 3120$, 2961, 1682, 1520, 1464, 1264, 1197 cm⁻¹; elemental analysis calcd $(\%)$ for $C_{105}H_{129}N_3O_{15}·7H_2O$: C 70.09, H 8.01, N 2.34, found: C 69.77, H 7.93, N 2.33.

Calix[6]cryptamide 10: The procedure described for the preparation of 4 (Method B) was applied to calix[6]trisamine 1 (100 mg, 0.0874 mmol) and compound 6 (42 mg, 0.0874 mmol). HBTU was used instead of PyBop and the reaction time was 15 h. The crude mixture was washed with an aqueous NaOH solution $(1 \text{ m}, 20 \text{ mL})$ and the flash chromatography eluent was CH_2Cl_2/a cetone (90:10). Calix[6]cryptamide 10 was isolated as a white solid (20 mg, 13%). The yield was calculated taking into account the presence of about 1 equiv of 1,1,3,3-tetramethylurea in the final product (1 H NMR estimation). 1 H NMR (300 MHz, CDCl₃): δ 0.89 (s, 27H; tBu), 1.25 (s, 27H; tBu), 2.24 (s, 9H; OMe), 3.34 (d, J=15 Hz, 6H; ArC H_{eq}), 3.72 (s_b, 6H; OCH₂CH₂N), 3.98 (s_b, 6H; OCH₂CH₂N), 4.14 (s, 6H; ArCH₂Ar), 4.37 (d, J=14 Hz, 6H; ArCH_{ax}), 6.67 (s_b, 3H; NHCO), 6.80 (s, 6H; ArH_{cal}), 7.05 (t, J = 7 Hz, 3H; ArH_{cap}), 7.11 (s, 6H; ArH_{cal}), 7.20-7.45 ppm (m, 12H; ArH_{cap}); HRMS (ESI-TOF): m/z calcd for C₁₀₅H₁₂₃N₃O₉ [M+Na]⁺: 1592.9191; found: 1592.9157.

Calix[6]cryptamide 11: The procedure described for the preparation of 4 (Method B) was applied to calix[6]trisamine 1 (100 mg, 0.0874 mmol) and compound 7 (46 mg, 0.0874 mmol). HBTU was used instead of PyBop and the reaction time was 15 h. The crude mixture was washed with an aqueous NaOH solution (1 M, 15 mL) and the flash chromatography eluent was CH_2Cl_2/a cetone (90:10). Calix[6]cryptamide 11 was isolated as a white solid (30 mg, 21%). M.p.: 200 °C (decomp); 1 H NMR $(300 \text{ MHz}, \text{CDCl}_3): \delta = 0.73 \text{ (s, 27H; } t\text{Bu}), 1.36 \text{ (s, 27H; } t\text{Bu}), 1.66 \text{ (s, 9H; }$ OMe), 3.38 (d, J = 15 Hz, 6 H; ArC H_{eq}), 4.12 (s_b, 6 H; OCH₂CH₂N), 4.15 $(s_b, 6H; OCH₂CH₂N), 4.58$ (d, $J=15 Hz, 6H; ArCH_{av}), 5.66$ (s, 6H; ArCH₂O), 6.59 (s, 6H; ArH_{cal}), 6.97 (d, $J=8$ Hz, 3H; ArH_{cap}), 7.07 (t, $J=8$ Hz, 3H; ArH_{cap}), 7.22 (s, 6H; ArH_{cal}), 7.38 (t, $J=8$ Hz, 3H; ArH_{cap}), 7.50 (s, 3H; ArH_{cap}), 8.28 (d, $J=8$ Hz, 3H; ArH_{cap}), 9.11 ppm $(s_b, 3H; NHCO);$ ¹³C NMR (75 MHz, CDCl₃): $\delta = 29.6, 31.2, 31.8, 34.2,$ 34.4, 40.7, 60.4, 69.4, 72.1, 112.8, 120.9, 121.7, 121.8, 123.7, 128.2, 132.7, 132.8, 133.2, 133.6, 138.9, 146.3, 146.4, 151.4, 154.5, 156.4, 165.9 ppm; IR (KBr): $\tilde{v} = 3700 - 3115$, 2961, 1661, 1599, 1528, 1483, 1298, 1210 cm⁻¹; HRMS (ESI-TOF): m/z calcd for C₁₀₅H₁₂₃N₃O₁₂ [M+Na]⁺: 1640.9004; found: 1640.9047.

Calix[6]cryptamide 12: The procedure described for the preparation of 4 (Method B) was applied to calix[6]trisamine 1 (100 mg, 0.0874 mmol) and CTV derivative 8 (66 mg, 0.0874 mmol). HBTU was used instead of PyBop and DMF instead of $CH₃NO₂$. The crude mixture was washed with an aqueous HCl solution $(1 \text{ m}, 15 \text{ mL})$ and the flash chromatography eluent was CH_2Cl_2/a cetone (55:45). After washing with diethyl ether, calix[6]cryptamide 12 was isolated as a white solid (53 mg, 33%). M.p.: 244 °C (decomp); ¹H NMR (300 MHz, CDCl₃/CD₃OD 3:1, 330 K): δ = 0.94 (s, 27H; tBu), 1.29 (s_b, 27H; tBu), 2.60 (s_b, 9H; OMe_{cal}), 3.32-3.70 (m, 6H; OCH₂CH₂N), 3.44 (d, J=14 Hz, 3H; ArCH_{eqcal}), 3.46 (d, J= 15 Hz, 3H; ArC H_{eucal}), 3.61 (d, J = 14 Hz, 3H; ArC H_{eacTV}), 3.74–4.02 (m, 12H; NHCH₂CO, OCH₂CH₂N), 3.95 (s, 9H; OMe_{CTV}), 4.42 (d, $J=14$ Hz, 3H; ArC H_{axcal}), 4.45 (d, J=15 Hz, 3H; ArC H_{axcal}), 4.58 (d, J=15 Hz, 3H; OCH₂CO), 4.67 (d, $J=15$ Hz, 3H; OCH₂CO), 4.77 (d, $J=14$ Hz, 3H; ArC H_{avCTV}), 6.79 (s, 3H; ArH_{cal}), 6.80 (s, 3H; ArH_{cal}), 6.96 (s, 3H; ArH_{CTV}), 7.00 (s, 3H; ArH_{CTV}), 7.12 ppm (s, 6H; ArH_{ca}); ¹³C NMR $(75 \text{ MHz}, \text{CDCl}_3)$: complex spectrum due to the presence of two conformations (see the text); IR (KBr): $\tilde{v} = 3725-3125$, 2960, 1663, 1513, 1481, 1265 cm⁻¹; HRMS (ESI-TOF): m/z calcd for C₁₁₁H₁₃₈N₆O₁₈ [M+Na]⁺: 1865.9965; found: 1865.9952.

Calix[6]cryptamide 13: The procedure described for the preparation of 4 (Method B) was applied to calix[6]trisamine 1 (100 mg, 0.0874 mmol) and calix[6]trisacid 9 (104 mg, 0.0874 mmol). The reaction time was 15 h and the flash chromatography eluent was $CH_2Cl_2/ACOE$ (98:2). Calix[6]-

cryptamide 13 was isolated as a white solid (117 mg, 58%). M.p.: 245° C (decomp); ¹H NMR (300 MHz, CDCl₃): $\delta = 0.82$ (s, 54 H; tBu), 1.37 (s, 27H; tBu), 1.39 (s, 27H; tBu), 2.45 (s, 9H; OMe), 2.48 (s, 9H; OMe), 3.44 (d, $J=16$ Hz, 6H; ArC H_{eq}), 3.45 (d, $J=15$ Hz, 6H; ArC H_{eq}), 3.86 $(s_b, 6H; OCH_2CH_2N), 4.04 (s_b, 6H; OCH_2CH_2N), 4.45 (s, 6H;$ OCH₂CO), 4.55 (d, $J=15$ Hz, 6H; ArC H_{ax}), 4.60 (d, $J=15$ Hz, 6H; ArCH_{ax}), 6.64 (s, 12H; ArH), 7.25 (s, 6H; ArH), 7.26 (s, 6H; ArH), 7.81 ppm (s_b, 3H; NHCO); ¹³C NMR (75 MHz, CDCl₃): δ = 30.3, 30.4, 31.3, 31.4, 31.8, 34.1(7), 34.2(2), 34.4, 34.5, 40.9, 53.6, 60.6(0), 60.6(1), 72.9, 123.9, 124.1, 128.1, 128.3, 132.8, 133.2, 133.3, 133.6, 146.0, 146.1, 146.3, 146.7, 152.5, 152.7, 154.9, 155.1, 169.9 ppm; IR (KBr): $\tilde{v} = 3715$ to 3090, 2962, 1682, 1481, 1201, 1122 cm-1 ; elemental analysis calcd (%) for C150H195N3O15·3H2O: C 77.18, H 8.68, N 1.80, found: C 77.09, H 8.67, N 1.76.

Host-guest complex 12 MI: IMI (2 equiv) was added to a solution of calix[6]cryptamide 12 (4.0 mg, 2.17 μ mol) in CDCl₃ (500 μ L) leading, after sonication, to the host-guest complex 12 MI as a unique species. ¹H NMR (300 MHz, CDCl₃, 263 K): $\delta = -0.10 - 0.14$ (m, 4H; CH_{2IMIin}), 0.70 (s, 27H; tBu), 1.37 (s, 27H; tBu), 3.34 (d, $J=15$ Hz, 3H; ArC H_{eqcal}), 3.42 (d, $J=15$ Hz, 3H; ArC H_{eqcal}), 3.35–3.45 (m, 6H; OCH₂CH₂N), 3.59 (d, J = 14 Hz, 3H; ArC H_{eqCTV}), 3.70 (s, 9H; OMe_{cal}), 3.80–4.10 (m, 12H; OCH₂CH₂N + NHCH₂CO), 3.94 (s, 9H; OMe_{CTV}), 4.35 (d, $J=15$ Hz, 3H; ArCH_{axcal}), 4.37 (d, J=15 Hz, 3H; ArCH_{axcal}), 4.60-4.72 (m, 6H; OCH₂CO), 4.77 (d, J = 14 Hz, 3H; ArC H_{avCTV}), 6.49 (s, 3H; ArH_{cal}), 6.55 $(s, 3H; ArH_{cal}), 6.86 (s, 3H; ArH_{CTV}), 7.05 (s, 3H; ArH_{CTV}), 7.24 (s, 3H;$ ArH_{cal}), 7.29 (s, 3H; ArH_{cal}), 8.05 (s_b, 3H; NHCO_{CTV}), 8.84 ppm (s_b, 3H; $NHCO...$).

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